

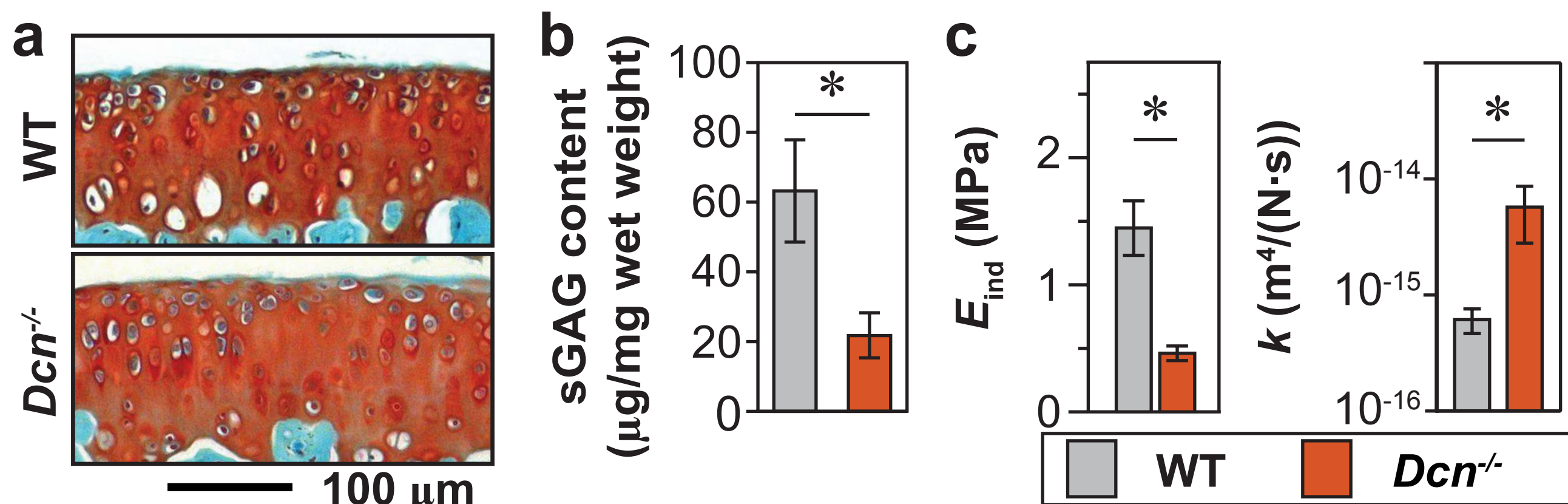
# Decorin Provides Molecular Linkages to Regulate the Integrity of Aggrecan Networks in Cartilage

## Objective

- To determine the role of **decorin**, the most abundant small leucine-rich proteoglycan (SLRP) in cartilage, in mediating the content and structural integrity of aggrecan in cartilage extracellular matrix (ECM).

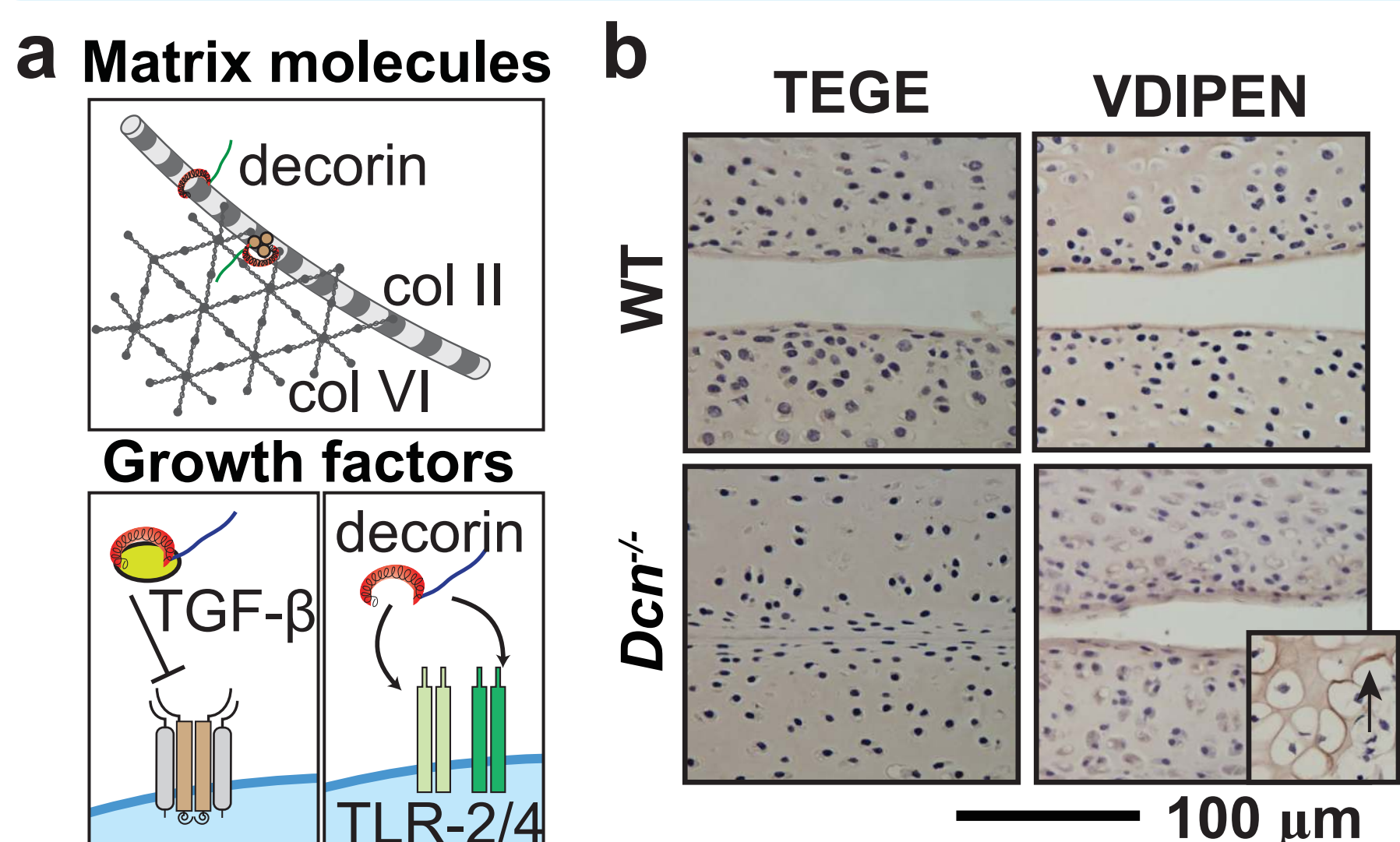
## Background

- The structural integrity of aggrecan, and its sulfated GAG side chains (sGAGs), are the key determinant of cartilage ECM mechanical functions [1].
- Our recent study showed that decorin knockout (*Dcn*<sup>-/-</sup>) murine cartilage develops substantially reduced aggrecan and sGAGs contents, resulting in impaired elastic and poroelastic mechanical properties (Fig. 1) [2].



**Fig. 1** Structure and mechanical phenotype of *Dcn*<sup>-/-</sup> mice, including a) reduced Safranin-O staining, b) substantial reduction in sGAG content, and c) lower indentation modulus and higher hydraulic permeability.

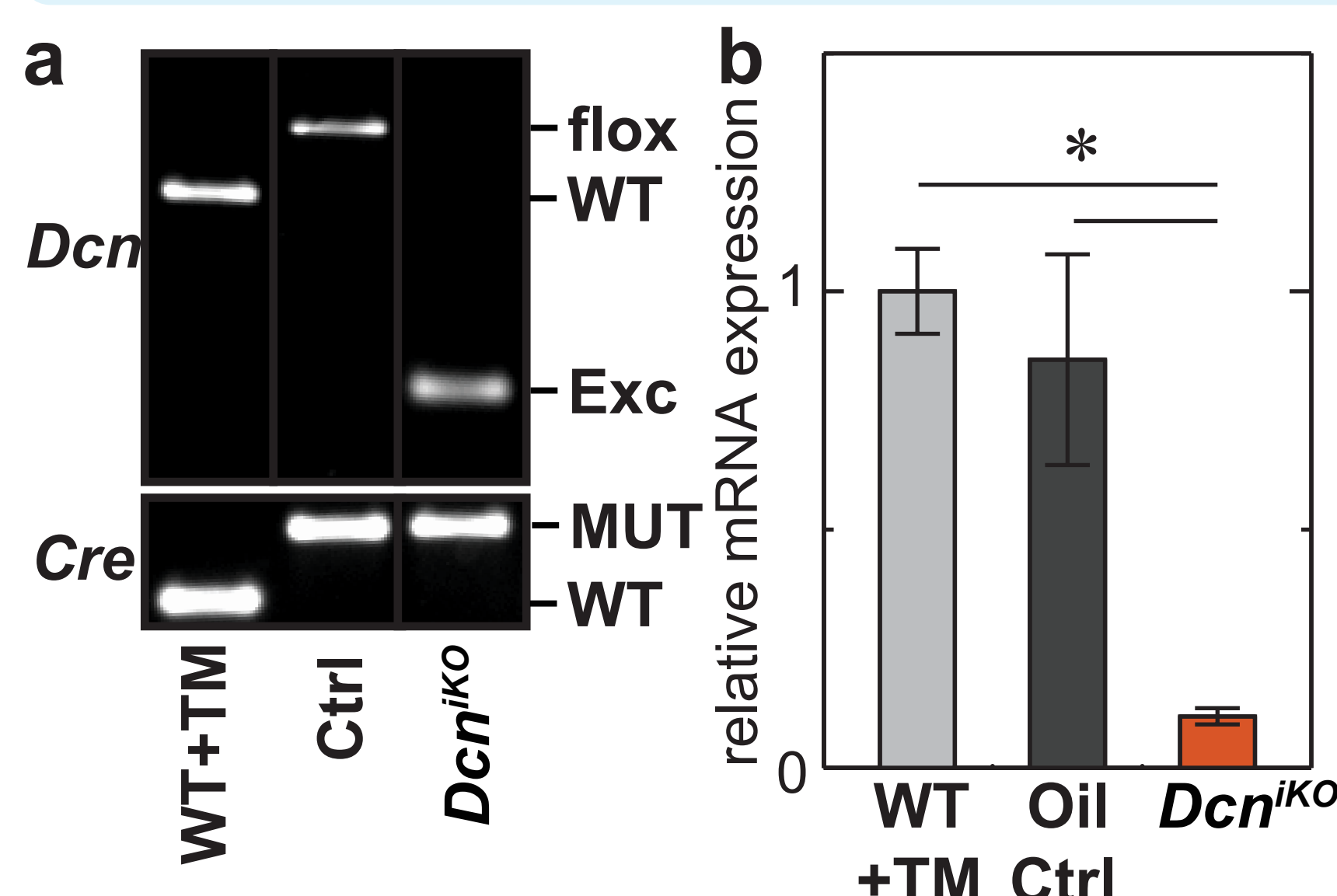
- Decorin interacts with a wide array of matrix molecules [3,4], growth factors and surface receptors [5-7] (Fig. 2a). This indicates that the aggrecan content phenotype could arise from factors including decrease in synthesis, increase in degradation and/or decrease in retention.
- In developing *Dcn*<sup>-/-</sup> cartilage, we did not detect degradation neo-epitopes of aggrecan by aggrecanases (TEGE<sub>373</sub>) [8] or MMPs (VDIPEN<sub>341</sub>) [9] (Fig. 2b), suggesting that decorin does not affect aggrecan degradation in vivo.
- Canonical functions of decorin include interaction with collagen [4] and aggrecan [3], and binding with TGF- $\beta$  to sequester TGF- $\beta$  signaling [10], indicating possible roles in mediating aggrecan synthesis and/or retention.



**Fig. 2** a) Schematics of decorin interactions with matrix molecules and growth factors. b) IHC shows no differences in the degradation neo-epitopes of aggrecan (Inset: positive control by black arrow).

## Materials and Methods

- Cell extraction and culture:** Primary chondrocytes were isolated from 1-month-old WT and *Dcn*<sup>-/-</sup> murine femoral head cartilage, and cultured in alginate beads. DMMB assay [11] was performed at 2, 4, 6 and 8 days of culture, and qPCR was performed on day 8.
- Decorin-aggrecan and decorin-collagen molecular interactions via AFM:** Gold-coated silicon substrates and microspherical AFM tips ( $R \approx 2.25 \mu\text{m}$ ) were end-functionalized with aggrecan [12]. Trypsin-treated bovine cartilage surfaces were used as the 2D collagen II network [13]. In PBS, with or without recombinant murine decorin protein, the tip was programmed to compress the substrate, held for 30 seconds, and then retracted at the same rate. The maximum adhesion force,  $F_{ad}$ , and total adhesion energy,  $E_{ad}$ , were calculated from each approach-retract force curve (e.g., Fig. 5a).
- Decorin inducible knockout mice:** Using *Dcn*<sup>fl/fl</sup>/*Rosa26CreER* (*Dcn*<sup>flKO</sup>) mice [14], we ablated the expression of decorin via tamoxifen injection (3 mg/40 g body weight) at 1-month age *Dcn*<sup>flKO</sup> (Fig. 3), and tested at 3-month age with Safranin-O/Fast Green histology and DMMB to assess joint morphology and sGAG content, and AFM-nanorheometric test [15] to quantify poroelastic mechanical properties. Control mice include the same iKO mice treated with vehicle, and WT mice injected with the same dose of tamoxifen.

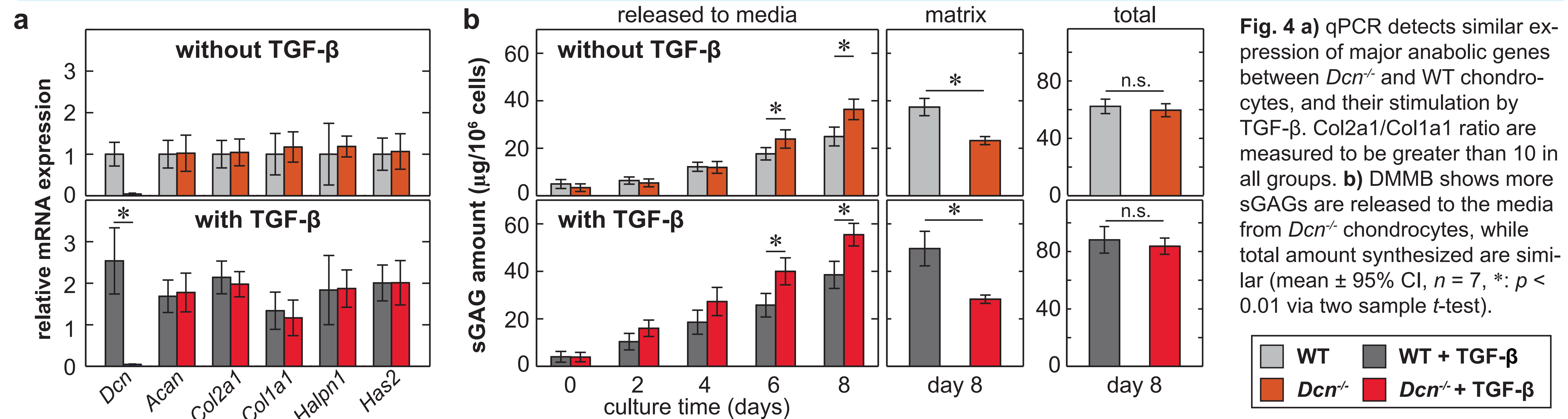


**Fig. 3** a) Genotyping of WT and *Dcn*<sup>flKO</sup> mice before and after tamoxifen injection showed the excision of floxed *Dcn* allele. b) qPCR showed daily injection of tamoxifen significantly reduced the expression of decorin to the baseline level on day 5 in cartilage.

## Results and Discussion

### 1) Decorin mediates the assembly and stability of aggrecan in neo-matrix, but does not affect its synthesis by chondrocytes.

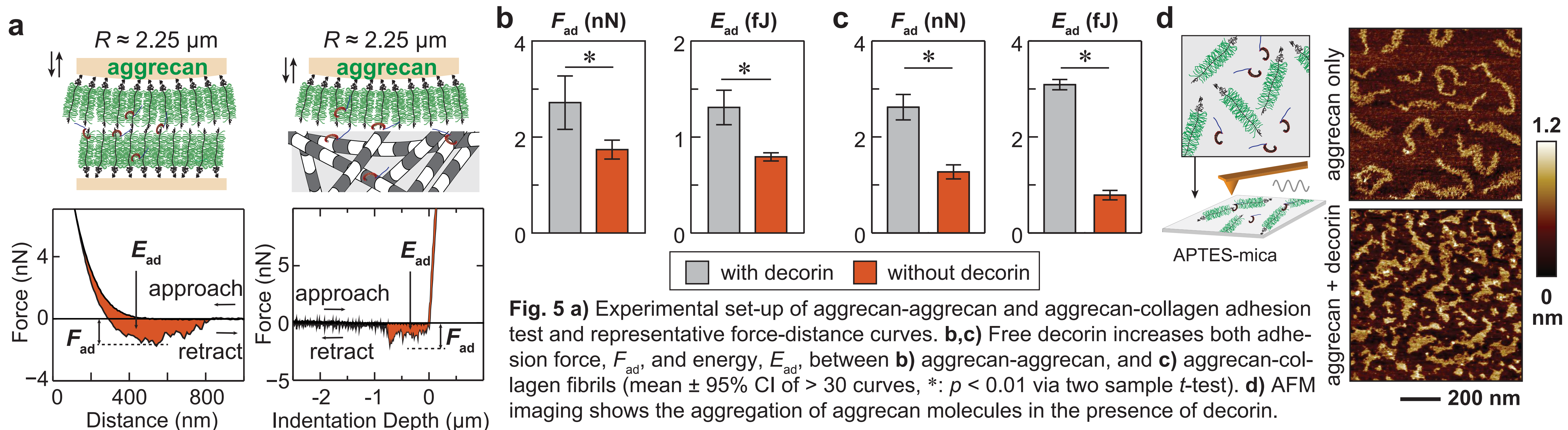
- Dcn*<sup>-/-</sup> chondrocytes exhibited similar expressions of matrix genes other than decorin, with or without TGF- $\beta$  (Fig. 4a). As expected, TGF- $\beta$  treatment enhances anabolism for both genotypes (Fig. 4a).
- In accordance with qPCR results, *Dcn*<sup>-/-</sup> chondrocytes synthesize similar total amount of sGAGs as WT, with or without the stimuli by TGF- $\beta$  (Fig. 4b).
- A significantly reduced proportion of sGAGs was retained in the neo-matrix of *Dcn*<sup>-/-</sup> chondrocytes, suggesting a key role of decorin in mediating aggrecan assembly, rather than synthesis (Fig. 4b).



**Fig. 4** a) qPCR detects similar expression of major anabolic genes between *Dcn*<sup>-/-</sup> and WT chondrocytes, and their stimulation by TGF- $\beta$ . Col2a1/Col1a1 ratio are measured to be greater than 10 in all groups. b) DMMB shows more sGAGs are released to the media from *Dcn*<sup>-/-</sup> chondrocytes, while total amount synthesized are similar (mean  $\pm$  95% CI,  $n = 7$ , \*:  $p < 0.01$  via two sample  $t$ -test).

### 2) Decorin increases aggrecan-aggrecan and aggrecan-collagen fibrils adhesion.

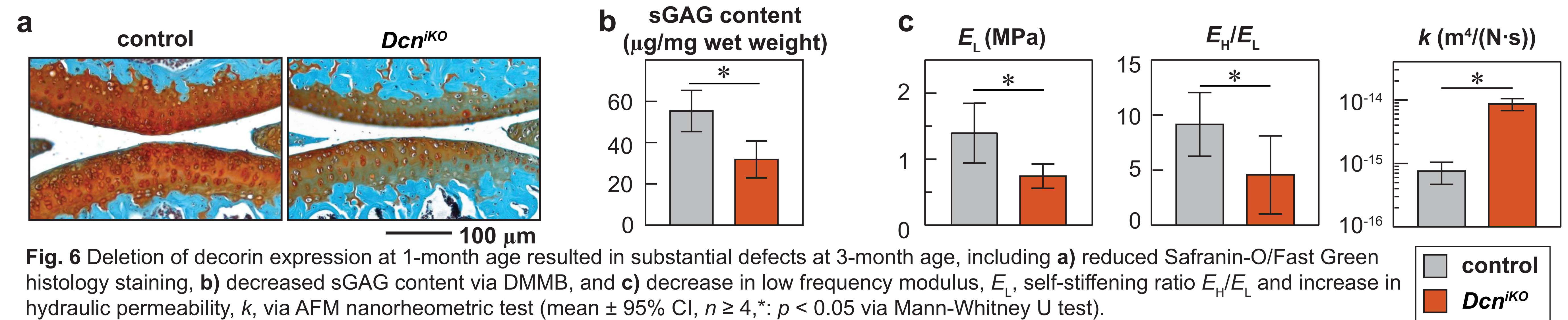
- Under AFM colloidal force spectroscopy (Fig. 5a) [12], adding free decorin significantly increased both the maximum adhesion force,  $F_{ad}$ , and adhesion energy,  $E_{ad}$ , both between aggrecan and aggrecan (Fig. 5b), and between aggrecan and collagen fibrils (Fig. 5c).
- When purified bovine aggrecan was reconstituted on atomically flat mica surface, adding free decorin transforms aggrecan monomers into interconnected supramolecular networks, shown by AFM imaging (Fig. 5d).



**Fig. 5** a) Experimental set-up of aggrecan-aggrecan and aggrecan-collagen adhesion test and representative force-distance curves. b,c) Free decorin increases both adhesion force,  $F_{ad}$ , and energy,  $E_{ad}$ , between b) aggrecan-aggrecan, and c) aggrecan-collagen fibrils (mean  $\pm$  95% CI of > 30 curves, \*:  $p < 0.01$  via two sample  $t$ -test). d) AFM imaging shows the aggregation of aggrecan molecules in the presence of decorin.

### 3) The role of decorin is manifested through late maturation, when collagen turnover is slow.

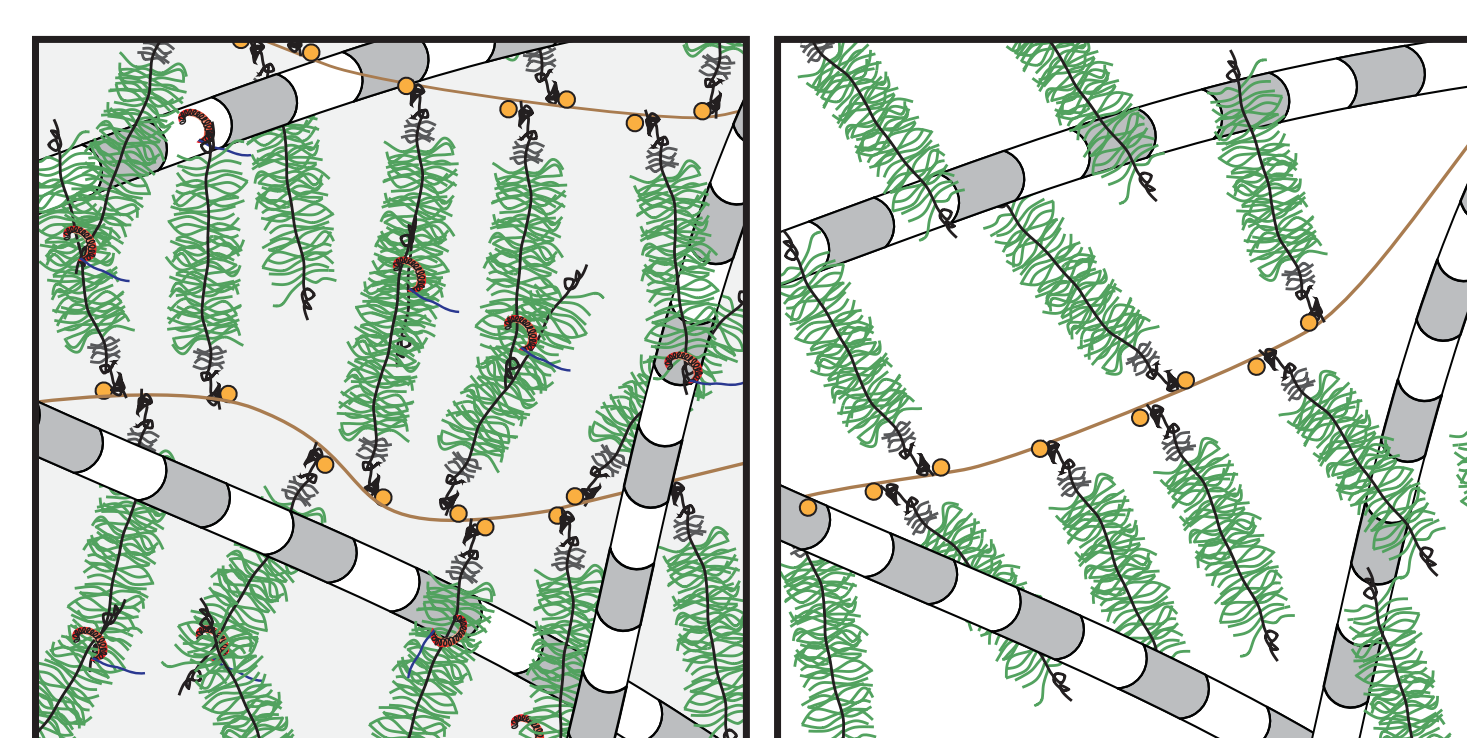
- After decorin was ablated in young adult mice (1-month age), similar structural and mechanical defects were observed in adult mice (3-month age) (Fig. 6).
- At the later stages of maturation, the turnover of collagen fibrils is slow [16], indicating the impact of decorin on aggrecan integrity is possibly independent of collagen fibrillogenesis in cartilage.



**Fig. 6** Deletion of decorin expression at 1-month age resulted in substantial defects at 3-month age, including a) reduced Safranin-O/Fast Green histology staining, b) decreased sGAG content via DMMB, and c) decrease in low frequency modulus,  $E_L$ , self-stiffening ratio  $E_H/E_L$  and increase in hydraulic permeability,  $k$ , via AFM nanorheometric test (mean  $\pm$  95% CI,  $n \geq 4$ , \*:  $p < 0.05$  via Mann-Whitney U test).

## Conclusion and Outlook

- Our results suggest that decorin increases aggrecan stability and retention via increasing its molecular adhesion, and this role is critical to the integrity of cartilage aggrecan network during post-natal growth (Fig. 7).**
- This new role of decorin is a clear departure from its known functions in mediating collagen fibrillogenesis, and thus, extends our understanding of decorin's structural role in various tissues.
- Building on these results, our ongoing studies are testing the potential of using decorin to improve regeneration by modulating the assembly of neo-matrix and mechanotransduction of chondrocytes.



**Fig. 7** Schematics show the hypothesized role of decorin in cartilage, and the impact of decorin loss on the reduction of aggrecan.

**References:** [1] Williamson +, J. Orthop. Res., 2001. [2] Han +, Trans. ORS, 2017. [3] Douglas +, Biomacromolecules, 2006. [4] Wiberg +, J. Biol. Chem., 2003. [5] Santra +, J. Biol. Chem., 2002. [6] Schönherr +, J. Biol. Chem., 2005. [7] Iozzo, Crit. Rev. Biochem. Mol. Biol., 1997. [8] Lark +, J. Biol. Chem., 1995. [9] Singer +, J. Clin. Invest., 1995. [10] Roughley +, Matrix Biol., 1994. [11] Farndale +, Biochimica. et biophysica acta, 1986. [12] Han +, Biophys. J., 2008. [13] Rojas +, Biomacromolecules, 2014. [14] Robinson +, Matrix Biol., 2017. [15] Nia +, J. Biomech., 2015. [16] Verzijl +, J. Biol. Chem., 2000.

**Acknowledgments:** This work was supported by the NIH (AR066824) and NSF (CMMI-1662544). We would like to thank Dr. David E. Birk for providing the breeding pairs of *Dcn*<sup>flKO</sup> mice and for valuable discussions. **Contact:** bh462@drexel.edu. **Disclosure:** The authors have no conflicts of interest.