

## Chapter

# Epigenetics of Circadian Rhythm Disruption in Cardiovascular Diseases

*Ivana Škrlec*

## Abstract

Circadian rhythm influences the regulation of homeostasis and physiological processes, and its disruption could lead to metabolic disorders and cardiovascular diseases (CVD). CVDs are still the dominant cause of death worldwide, which are related to numerous environmental and hereditary risk factors. Environmental and hereditary factors can clarify a small fraction of the CVD risk discrepancy. Epigenomics is a very bright strategy that will complement the knowledge of the genetic basis of CVDs. Epigenetic mechanisms allow cells to reply promptly to environmental changes and include DNA methylation, histone modification, and non-coding RNA alterations. According to research data, the circadian rhythm regulates many epigenetic regulators. The challenge is to understand how epigenetic events happen rhythmically in tissues that are involved in the development of CVDs. Epigenetic events are possibly reversible through their interface with environmental and nutritional factors, allowing innovative preventive and therapeutic strategies in cardiovascular diseases.

**Keywords:** circadian rhythm, cardiovascular disease, epigenetics, DNA methylation, histone modification, microRNA

## 1. Introduction

The word epigenetics comes from the Greek word ‘epi’ that means above; that is, hereditary variations in phenotype that do exclude alterations in the nucleotide sequence in DNA [1]. Epigenetic mechanisms involve DNA methylation, post-translational histone modifications, and noncoding RNAs (ncRNAs) [1]. Many studies focus on the epigenetic mechanisms of various diseases. Epigenetic processes are essential for the healthy growth and development of an organism [1]. Epigenetic mechanisms are implicated in the expression of circadian genes in the suprachiasmatic nucleus (SCN) neurons and peripheral tissues [2]. The accumulation of lifestyle and age-related epigenetic changes could result in the development of metabolic disorders and atherosclerosis [2].

The influence of epigenetic changes on the cardiovascular system is an essential link between genotype to phenotype diversity [3]. Epigenetic changes are potentially reversible and may be affected by environmental factors, nutrition, as well as gene-environment interactions. Identifying and understanding epigenetic factors represent a new insight into our knowledge of the risks of cardiovascular disease (CVD) [1].

## **1.1 Circadian rhythm**

The circadian clock is a preserved system that allows organisms to adapt to frequent daily variations, such as the day and night and food availability [4]. This center clock receives signals from the environment and coordinates the daily activity of peripheral clocks found in almost all tissues [4]. The molecular clock is vital in maintaining metabolic and physiological homeostasis [5]. The circadian clock is linked to cellular metabolism so that dysregulation of the circadian rhythm can contribute to various pathological conditions such as diabetes, obesity, metabolic syndrome, inflammation, sleep disorders, and CVDs [5–8].

Genome-wide studies show that 10–15% of all transcripts have a circadian pattern in different tissues involved in the control of metabolism, such as the cardiovascular function [4, 6, 8, 9]. The onset of ischemic cardiopathy is irregularly distributed during the day [1, 10, 11]. A chronobiological strategy to heart disease may present new possibilities to enhance drug development to improve therapeutic outcomes [1]. Genetic evidence supports the function of circadian rhythm in the adjustment of metabolism.

## **1.2 Cardiovascular diseases**

Cardiovascular diseases are complex and diverse. They include hypertension, coronary artery disease, heart failure, and stroke and are a main worldwide reason for morbidity and death in advanced economies and carry a substantial economic burden [1, 3, 12–15]. CVDs are associated with a variety of hereditary and variable risk factors, but environmental and genetic impacts may explain a smaller fraction of CVD risk variability [1, 12]. Studies showed that there is a wide range between 40 and 80% of the genetic contribution to the onset of cardiovascular disease [16].

The complex pathogenesis of CVD is due to the abundance of genetic and environmental factors, of which epigenetic changes are a significant factor [3]. Several risk factors of CVD, like diet, smoking, stress, circadian rhythm, and pollution, are related to epigenetic modifications [1]. Disorders such as hypertension, diabetes, and obesity are often utilized to recognize and cure people at increased CVDs risk [1]. Epigenetic modifications are associated with the processes involved in the CVD in humans or directly affect the gene expression involved in a major cardiac complication, myocardial infarction (MI). Hypertension is one of the leading causes of CVDs [3], while insulin resistance is one of the most significant precursors of type 2 diabetes and associated cardiometabolic conditions [17].

Changes in the style of living and diet could decrease the risk of CVDs [14]. Epigenetic factors indicate there is interindividual variability from birth. It can be stable over the life span and is considered to be an initiator of early programming for adult-onset diseases [12, 18]. The understanding of epigenetics in the onset of CVDs may provide a new perspective on diseases [14].

## **1.3 Epigenetics**

Epigenetics studies heritable variations in gene expression that exclude any change in the DNA sequence [16, 19]. Epigenetic changes include modifications of the DNA base, post-translational histone modifications, and ncRNA mechanisms that run in the nucleus [16, 20]. The epigenome moves the genome from a transcriptionally active to a transcriptionally inactive state [4, 21]. Epimutation transmissions occur throughout the life of the individual [2]. The rate of epigenetic variation is higher than that of genetic mutations because the formation of new inherited changes allows adjustment to a new environment [14, 16].

The most studied epigenetic change is cytosine methylation. It is also a method for suppressing gene expression [22]. DNA methyltransferase (DNMT) enzymes perform DNA methylation. DNMTs bind the methyl group to the 5-site cytosine [16]. The methyl group most commonly binds to the cytosine at a CpG site. It is the fundamental and ubiquitous epigenetic mechanism [14]. The DNMT enzyme family, consisting of DNMT1, DNMT3a, and DNMT3b, methylates cytosine into 5-methylcytosine [14]. Promoter methylation is usually connected with inhibition of transcription [14]. DNMT1 controls the mitotic inheritance of methylated DNA, while DNMT3a and DNMT3b are mainly in charge of *de novo* methylation [14]. The different epigenetic modification is DNA hydroxymethylation, including 5-hydroxymethylated cytosines [14]. Different nutritional and lifestyle factors can affect the methylation of particular CpG sites in gene promoters and into adulthood [22].

Nucleosomes are composed of histone proteins around which DNA is wound into chromatin [16]. Nucleosomes consist of eight histone proteins: two dimers of H2A/H2B and two dimers of H3/H4. Each histone has an adjustable amino-acid tail [16]. Histones can change at more than 30 amino acid residues within amino-terminal tails [4]. Histone modifications include various processes such as acetylation, methylation, phosphorylation, sumoylation, and ubiquitination. It has a function in the organization of chromatin composition and gene expression by altering the intensity of chromatin condensation [1, 14, 23]. Histones are mostly acetylated on lysine (K) residues. Histone acetyltransferase (HAT) and histone deacetylase (HDAC) regulate histone acetylation [14]. Histone methyltransferase regulates histone methylation, while histone demethylase catalyzes demethylation. Transcription activation is usually associated with acetylation of lysine residues at histones 3 (H3) and 4 (H4). Depending on the location of the target lysines in the histone tail and the number of methyl groups added, methylation can either activate or inhibit gene expression [14, 24]. Histone phosphorylation is a marker of cell division and has a function in DNA repair, chromatin condensation during division, and regulation of gene expression [14, 25]. The addition of ubiquitin to lysine residues in histones is called ubiquitination and is implicated in DNA repair and control of transcription [14]. Sumoylation is a changeable post-translational adjustment using small ubiquitin-like proteins (SUMO) and has a crucial function in various mechanisms, such as transcription, and cell cycle progression [14, 26].

RNA-based epigenetic mechanisms include long noncoding RNAs (lncRNAs) and microRNAs (miRNAs) [14]. ncRNAs are functional RNAs that do not translate into proteins and play an essential part in epigenetic regulation [14, 16]. The lncRNAs are extremely tissue-specific relative to protein-coding genes [16]. The miRNAs are short (20–22 nucleotides), single-stranded, evolutionarily-conserved ncRNAs that modulate the expression at the post-transcriptional level of more than 50% of cellular genes [13, 14, 27].

Changes in the environment, including temperature, light, and nutritional habits, trigger reversible epigenomic modification that can influence numerous physiological processes [28]. Epigenome-wide association studies (EWAS) provide information about associations between epigenomic perturbations and traits associated with human diseases [29]. EWAS try to evaluate the environmental impact on genetic regulation. The epigenetic variations could explain missing parts of heritability of chronic diseases that have not yet been determined by genome-wide association studies [29].

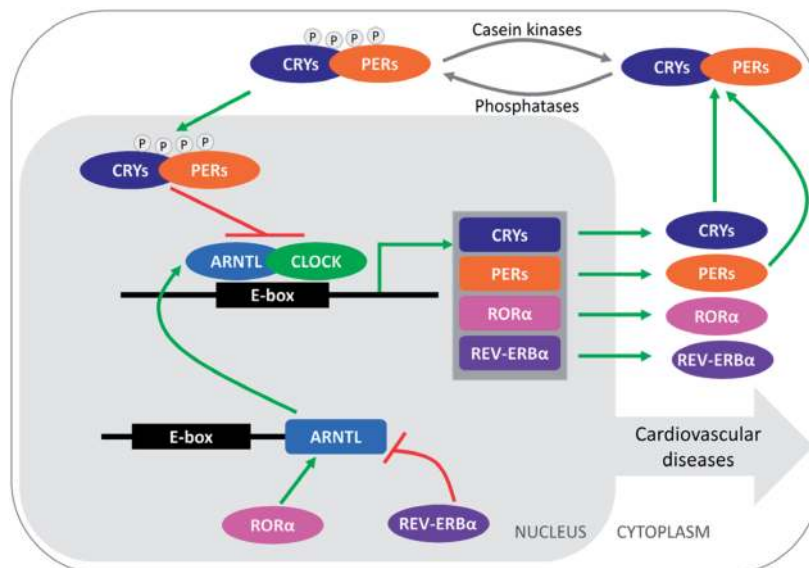
## **2. Molecular background of circadian rhythm**

The primary clock genes show circadian expression in the SCN, and light is one of the key drivers that can reset the rhythm phases. There are several crucial proteins

in SCN. Transcription activators are aryl hydrocarbon receptor nuclear translocator-like (ARNTL or BMAL1) and circadian locomotor output cycle caps (CLOCK). Transcription inhibitors are period (PER) and cryptochrome (CRY) [30, 31]. Within 24 h, the entire process of activation and inhibition of gene expression takes place [32, 33]. The circadian system controls gene expression through various mechanisms as a basis of global gene regulation. The first is via E-boxes (promoter and enhancer regulatory elements) of oscillator proteins such as CLOCK, ARNTL, and NPAS2 (neuronal PAS domain protein 2). The second mechanism is using other oscillator proteins such as ROR $\alpha$  (retinoic acid receptor-related orphan receptor) and REV-ERB $\alpha$  (or NR1D1, orphan nuclear receptor) via REV-ERB/ROR response element (RRE), which are present in the promoters of specific clock-controlled genes (CCGs) (Figure 1). The third mechanism is the daily chromatin remodeling [2, 19, 34, 35].

The ARNTL-CLOCK heterodimers enhance *CRY* and *PER* expression, as well as the expression of additional CCGs. Phosphorylated CRY-PER heterodimers repress the action of ARNTL-CLOCK heterodimer. As a result, *CRY* and *PER* gene transcription is decreased during the day, while ubiquitin degradation reduces the CRY and PER protein levels. The PER2 has histone deacetylase activity and modified chromatin structure, followed by transcription termination [36–39]. The new cycle begins with the termination of the ARNTL-CLOCK repression during the day. Casein kinase 1 (CK1) regulates the quantity of CRY-PER heterodimers' phosphorylation or degradation. CK1 controls protein activity via its phosphorylation [40]. An additional negative loop is REV-ERB $\alpha$ . It binds to the RRE of the *ARNTL* and *CLOCK* genes and inhibits their transcription. Overnight, REV-ERB $\alpha$  degrades, and ROR $\alpha$  elevates the *ARNTL* gene transcription [2, 32, 41]. ARNTL-CLOCK heterodimers increase transcription of the nuclear receptors ROR $\alpha$  and REV-ERB $\alpha$  and form an additional circadian rhythm loop [31, 42].

Nearly 10% of the transcripts show circadian rhythmicity [19]. Rhythmic expression of crucial metabolic genes is impaired due to clock gene mutations and lead to metabolic disorders [28]. Fasting glucose levels decrease, and insulin sensitivity



**Figure 1.**

*Circadian rhythm gene regulation in cardiovascular diseases. ARNTL and CLOCK activate transcription of CRY and PER and nuclear receptors (REV-ERB $\alpha$  and ROR $\alpha$ ). CRY and PER heterodimerize and phosphorylate by casein kinases and translate into the nucleus where they prevent binding of the ARNTL-CLOCK to the regulatory regions of target genes. In the second feedback loop, REV-ERB $\alpha$  prevents the transcription of ARNTL, while overnight the ROR $\alpha$  activates transcription of ARNTL. ARNTL—aryl hydrocarbon receptor nuclear translocator-like, CLOCK—circadian locomotor output cycle kaput, CRY—cryptochrome, PER—period, P—phosphate, ROR $\alpha$ —retinoic-related orphan receptor alpha, Ub—ubiquitin.*

increases in overexpression of the *CRY1* [28]. *ARNTL* deletion and *CLOCK* mutation disturb lipid metabolism [28, 43]. REV-ERB $\alpha$  is involved in liver circadian lipid biosynthesis, and REV-ERB $\alpha$  and *ARNTL* manage adipocyte differentiation [28]. A primary regulator of bile acid synthesis is REV-ERB $\alpha$ , while the *PER1* and *PER2* deletion upregulates bile acid biosynthesis and causes hepatic cholestasis [28].

Based on circadian rhythms in SCN neurons and peripheral cells, epigenetic mechanisms participate in the formation of circadian rhythms of gene expression [2]. One of the primary circadian genes, *CLOCK*, has the function of histone acetyltransferase. Chromatin remodeling is an essential underlying mechanism of the clock rhythm and reveals an association between cellular physiology and histone acetylation [2]. *ARNTL*-*CLOCK* heterodimer or *ARNTL*-*NPAS2* complex mobilizes HATs and HDACs [28, 44]. To maintain metabolic homeostasis and avoid metabolic disorders, the crosstalk between circadian rhythm and metabolism is necessary [28].

### 3. Epigenetic changes in circadian rhythm in cardiovascular diseases

Rapid adaptation of cells to environmental changes is facilitated by epigenetic mechanisms that also offer a link between genes and the environment [1]. The phenotypic variations observed in humans are more significant than genotype variations alone, and changes in epigenetic gene modification explain them [1, 45]. CVDs, such as atherosclerosis, cardiac hypertrophy, myocardial infarction, and heart failure, are associated with epigenetic mechanisms ranging from DNA methylation, histone modification, to ncRNAs [13]. An essential way of developing CVD early in life involves epigenetic changes [12]. The underlying mechanism providing the link between the early life environment and the subsequent CVD risk is epigenetic modifications [12].

The association of methylation with specific genes may be useful in assessing the risk of a disease or in monitoring the response to a particular treatment [14]. In the process of DNA methylation, homocysteine, an amino acid that does not enter into protein composition, is essential [46]. The lack of folate in the diet leads to an increase in plasma homocysteine, which contributes to the rise of S-adenosyl homocysteine. It represses transmethylation reactions and decreases methylation all over the epigenome [1, 46]. In atherogenesis are included homocysteine-induced changes in DNA methylation in smooth muscle vascular cells [1, 47, 48]. Endothelial dysfunction and different aspects of CVD are epigenetically associated with folic acid deficiency [16]. Genomic DNA is hypomethylated in human atherosclerotic lesions [1, 2, 12]. Inflammatory processes involved in the development of atherosclerotic plaques are associated with hypermethylation [1, 49]. There are rhythmic changes in global DNA methylation in human blood, and there is an increased level at night [35]. Changes in circadian rhythm genes methylation were observed in aging mice, but are tissue-dependent [35, 50]. For example, in the stomach of older mice, the methylation of the *PER1* promoter decreased, while the methylation of the *ARNTL*, *CRY1*, and *NPAS2* promoters in the spleen was increased [35]. Sleep disorders affect circadian rhythm gene methylation, especially *ARNTL*, *CRY1*, and *PER1* [35, 51]. Temporary epigenetic changes linked with rhythmic gene expression lead to circadian epiphenotypes [2]. Based on this, it can be concluded that DNA methylation may be reversed by conventional drugs, independent of DNA replication [2].

The histone code is involved in many aspects of cardiovascular physiology, from endothelial cell responses to hypoxia to recovery from MI [16]. *CLOCK* has enzymatic properties of histone acetyltransferase (HAT). It performs acetylation at Lys537 of H3 histone and *ARNTL*, which is necessary for circadian rhythm [1, 9]. *CLOCK* works in collaboration with other HATs to maintain circadian rhythm in the acetylation state of histones at CCG promoters [6]. HDAC activity has an essential function in defining

the intensity of myocardial ischemia, especially after MI [16]. Inhibition of HDAC can promote angiogenesis and reduce myocardial damage after MI [16], such as valproic acid (VPA), which is an HDAC inhibitor [2]. Histone deacetylases, SIRT1 (sirtuin 1), and SIRT6 participate in the histone modification, thus controlling gene expression [35] and providing a molecular connection among metabolism and circadian rhythm [6]. SIRT1 deacetylates regulatory proteins and acts as a rhythm-promoting agent in circadian oscillators [35]. SIRT1 has a unique role in central and peripheral circadian rhythms [35]. The purpose of histone phosphorylation in CVDs is minimal [14], while SUMO proteins influence the activity of several essential factors that are important for cardiac development [14]. There are connections between circadian rhythm regulators, chromatin modifications, and cellular metabolism [1, 52].

Numerous lncRNAs have essential regulatory functions in various CVDs [14]. The miRNAs regulate cholesterol metabolism, oxidative stress, and endothelial dysfunction, diverse cellular processes involved in atherosclerosis [14]. miRNAs may be relevant regulators of circadian rhythm [1]. Circulating miRNA-145 and miRNA-126 are decreased in patients with coronary artery disease, while miRNA-1, miRNA-499, and miRNA-133b are increased during acute myocardial infarction [13]. All those miRNAs can be biomarkers of CVD.

Circadian rhythms combine metabolic and environmental signals and alter gene expression when adapting the organism to particular circumstances [6]. Many epigenetic regulators in some tissues are controlled in a circadian fashion [19, 53]. The challenge is to determine whether epigenetic variations happen in a rhythmic pattern in tissues included in the CVD development [12, 19]. Epigenetics can contribute to enhancing CVD therapies and finding new markers for CVD screening [16, 54].

#### 4. DNA methylation and CVDs

DNA methylation is a durable, relatively constant epigenetic change. It involves the covalent attachment of a methyl group to the cytosine [3, 55]. The primary role of DNA methylation is to regulate gene expression by altering the availability of DNA to the transcription factors [3, 13].

DNA methylation links the steady genome and the changing environment. It is an instrument through which environmental changes influence metabolism [7]. Disruption of DNA methylation has been associated with different metabolic diseases such as diabetes [56], obesity, and insulin resistance [57]. Furthermore, the epigenetic mechanisms control circadian rhythm, and circadian disturbance leads to DNA methylation changes of the clock genes [7, 51, 58]. Adiposity, metabolic syndrome, and weight loss are linked to DNA methylation changes of the *ARNTL*, *CLOCK*, and *PER2* gene promoters [7]. It indicates the significance of determining the impact of DNA methylation in epigenetic studies in complex human disorders [7].

DNA methylation is cell- or tissue-specific, but epimutations are not restricted to the affected tissue and may also be observed in peripheral blood [7]. Compared to other genes, the regulatory regions of circadian rhythm genes are plentiful in CpG sites [59, 60]. Patients with coronary artery disease have altered methylation patterns relative to controls [1, 61, 62]. All mentioned supports the assumption that epigenetic variations are associated with an increased CVD risk [1].

Epigenetic alterations of circadian genes are related to obesity and metabolic disorders [17, 63]. A positive association was found between the alteration of the *ARNTL* gene methylation and weight loss, and its activity is included in the control of adipogenesis and lipid metabolism [63]. The long-term shiftwork, associated with obesity and metabolic syndrome risk, induces hypomethylation of the *CLOCK* gene promoter [22]. *CLOCK* gene SNPs are associated with a

predisposition to metabolic syndrome [22, 64]. Long-term shift work, obesity, and metabolic syndrome are associated with *CRY2* hypermethylation in peripheral blood [22]. Genetic variants of the human *PER2* gene are related to abdominal obesity and CVDs [22, 65]. The methylation status of CpG sites in the *PER2* gene is associated with obesity, metabolic syndrome, and weight loss [7, 22].

Global hypomethylation of DNA is present in atherosclerotic lesions [3, 66]. The severity of atherosclerotic lesions correlates with DNA methylation [3, 14, 67, 68]. There are notable variations in DNA methylation after an MI event [69, 70]. DNA methylation status in blood samples is related to CVD [71, 72].

Environmental and behavioral factors, such as inflammation, smoking, physical activity, or stress, can alter the epigenome [63, 73]. Elevated gene expression in the inflammatory pathway is associated with decreased gene methylation [46]. DNA methylation relies on the accessibility of methyl groups obtained from methionine, and the existence of certain nutrients in the food influences epigenetic changes with possible cardiovascular outcomes [46]. Although methylation changes are related to healthy aging, they could be in the background of the development of some diseases, such as CVD [46, 74]. Reduction in global DNA methylation occurs throughout the human lifespan [46].

## 5. Histone modification and CVDs

Post-translational modifications occur at amino acid residues in the amino-terminal regions of histone and cover histone acetylation, methylation, phosphorylation, sumoylation, and ubiquitination. It controls chromatin remodeling and gene expression [3, 23]. Histone acetylation is a sign of transcription activation [75], while histone methylation can both stimulate and inhibit transcription [28, 75]. Post-translational histone modifications control genes coding clock proteins [46, 75]. Epigenetic irregularities are related to different disorders, including atherosclerosis [4, 76].

Histone modifications occur at the CCG promoters in a circadian fashion [4, 44, 77, 78]. The core clock protein, CLOCK, has HAT activity. It revealed the molecular association among epigenetic mechanisms and circadian rhythm [4, 19, 59, 79]. CLOCK acetylates ARNTL, which facilitates CRY-dependent repression [19, 28], and interaction of CRY1 with the ARNTL-CLOCK heterodimer [9]. CLOCK and NPAS2 attract different HATs to the promoter of the *PER1* in vascular tissues [59, 78]. The rhythmic binding of ARNTL and CLOCK transcriptional activators directly influences the acetylation of specific histone lysine residues near the DNA-binding site without the involvement of additional HAT enzymes [59]. CLOCK acetylates additional non-histone proteins that have crucial roles in the regulation of different cellular events [4].

SIRT1 is an NAD<sup>+</sup>-dependent histone deacetylase [4, 59, 80]. It is needed for rhythmic transcription of some clock genes, such as *ARNTL*, *CRY1*, and *PER2* [80, 81]. SIRT1 represents the molecular connection between metabolic processes, chromatin remodeling, and circadian physiology [4]. SIRT1 plays a crucial role in metabolism. It deacetylates some proteins of the metabolic pathways and regulates gene expression by histone deacetylation [75]. *SIRT1* expression levels are nearly constant over 24 h, just like relatively constant *CLOCK* gene expression levels\*\*\* [4, 82–85]. The HAT function of CLOCK is balanced by SIRT1, which deacetylates H3 and ARNTL, and PER2 [79, 83, 86]. SIRT1 binds to ARNTL-CLOCK within a chromatin complex that, in a circadian fashion, binds to CCG promoters [4, 87]. ARNTL and PER2 are SIRT1 targets [4]. SIRT1 associates with ARNTL-CLOCK heterodimers and improves the deacetylation and degradation of PER2 [86]. SIRT1 deacetylates clock proteins in a circadian fashion [4]. HDAC3 is a deacetylase that modulates histone acetylation of circadian genes, especially those included in lipid

metabolism, such as REV-ERB $\alpha$  [88–92]. Mutations of circadian rhythm proteins that can either modify histones (such as CLOCK) or link to histone modifiers (such as ARNTL, PER2, and REV-ERB $\alpha$ ) are related to metabolic syndrome [75, 79]. Endogenous SIRT1 plays a crucial role in mediating cell death/survival processes and is involved in the pathogenesis of the CVDs [28, 93]. The ARNTL sumoylation plays an essential role in ARNTL accumulation and circadian rhythmicity [86].

Histone modifications, and particularly HDACs, have a significant role in the control of vascular homeostasis. Dysregulation of HDAC could lead to the formation of atherosclerotic lesions [14, 94]. In human carotid arteries, histone methylation and acetylation present recognizable patterns depending on the seriousness of the plaque [46]. Inhibition of HDACs leads to reduced inflammation and atherogenesis [46, 95]. In animal studies, HDAC inhibitors reduce the size of MI and ischemia-reperfusion injury after revascularization [46, 96]. The inhibition of HDAC may improve myocardial recovery and block post-infarction remodeling [46]. Fibrosis after MI was reduced by valproic acid, an HDAC inhibitor [14, 97].

## **6. MicroRNAs and CVDs**

MicroRNAs (miRNAs) are small noncoding RNA molecules that repress the expression of target messenger RNAs [1, 3, 5, 98]. MicroRNA dysregulation is associated with cardiovascular diseases, lipid metabolism, endothelial function, ventricular hypertrophy, and post-infarction dysrhythmias [1, 5].

Oscillating microRNAs, based on external triggers, could affect the expression of target genes in a circadian fashion independently of clock genes [5, 99]. In plasma and serum of CVD patients are observed decreased levels of numerous miRNAs, such as miRNA-126, miRNA-17, miRNA-145, miRNA-92a, and miRNA-155 [3].

MiRNAs control the development of atherosclerosis through their action on endothelial function, plaque progression and rupture, and blood vessel development [46]. MiRNA-126 expressed by endothelial cells serves as an adverse adjuster of vascular inflammation, while miRNA-33 plays a vital role in inhibiting the critical genes implicated in cellular cholesterol export [14, 100]. Some miRNAs target DNMTs and thus regulate the level of DNA methylation in atherosclerotic lesions [14]. MiRNA-148 changes HDL and LDL cholesterol levels in murine models and thus has a vital function in lipid metabolism [46, 101, 102].

MiRNA-24, 29a, and 30a influence the circadian rhythm by regulating the stability and translation of PER1 and PER2 mRNAs [5]. The ARNTL-CLOCK heterodimer controls miRNA-142–3p and, in turn, can target ARNTL [5, 103, 104]. MiRNA-21 is a PER2-dependent miRNA and mediates PER2-obtained cardioprotection [5, 105]. Through cellular stress, PER2-dependent miRNA-21 controls cellular glycolysis. Myocardial ischemia causes activation of pathways aimed at increasing the efficiency of myocardial oxygen [5, 106]. Suppression of miRNA-21 reduces the fibrotic response and enhances cardiac activity [5].

A valuable sign of myocardial cell death is the plasma levels of miRNA-208 [3, 107]. MiRNAs have a function in remodeling after MI, a mechanism closely associated with the expansion of tissue fibrosis [14]. A more sensitive biomarker of acute non-STEMI is miRNA-499 than cardiac troponin T [46].

MicroRNAs could potentially become new modulators of circadian rhythms and could have a positive effect on cardiovascular physiology [5]. MiRNAs regulate about 60% of all human genes [46]. Therapeutic strategies should target specific microRNAs and thus reduce their capacity to inhibit circadian rhythm components or circadian rhythm output genes [5, 108–112]. Administration of microRNAs in a circadian-dependent fashion could serve to adapt the impaired circadian system,



advance metabolism by enhancing efficient oxygen pathways, and thereby promote cardioprotection from ischemia [5].

## 7. Conclusion

The epigenetic variations of an individual change throughout a lifetime and epigenome profiles, instead of genotypes, are reflected in phenotypes in epigenetic epidemiological studies. Therefore, epigenetic modifications are the reason or a result of a pathological condition. Understanding the epigenetic contribution to CVD pathology may help to develop new treatments and diagnostic approaches. Epigenetic biomarkers might be very useful in treatment monitoring and predicting disease outcome. Epigenetic events can potentially be reversibly altered depending on environmental and nutritional factors. Understanding epigenetic mechanisms may identify valuable, novel biomarkers for disease.

## Conflict of interest

The author declares no conflict of interest.


## Author details

Ivana Škrlec  
Histology, Genetics, Cellular, and Molecular Biology Laboratory, Faculty of Dental Medicine and Health, Department of Biology and Chemistry, J.J. Strossmayer University of Osijek, Croatia

\*Address all correspondence to: [iskrlec@fdmz.hr](mailto:iskrlec@fdmz.hr)

## IntechOpen

---

© 2020 The Author(s). Licensee IntechOpen. This chapter is distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/3.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited. 

## References

- [1] Ordovás J, Smith C. Epigenetics and cardiovascular disease. *Nature Reviews. Cardiology*. 2011;7:510-519. DOI: 10.1038/nrcardio.2010.104.Epigenetics
- [2] Gallou-Kabani C, Vigé A, Junien C. Lifelong circadian and epigenetic drifts in metabolic syndrome. *Epigenetics*. 2007;2:137-146. DOI: 10.4161/epi.2.3.4897
- [3] Udali S, Guarini P, Moruzzi S, Choi S-W, Friso S. Cardiovascular epigenetics: From DNA methylation to microRNAs. *Molecular Aspects of Medicine*. 2013;34:883-901. DOI: 10.1016/j.mam.2012.08.001
- [4] Bellet MM, Sassone-Corsi P. Mammalian circadian clock and metabolism—The epigenetic link. *Journal of Cell Science*. 2010;123:3837-3848. DOI: 10.1242/jcs.051649
- [5] Oyama Y, Bartman CM, Gile J, Eckle T. Circadian microRNAs in cardioprotection. *Current Pharmaceutical Design*. 2017;23:3723-3730. DOI: 10.2174/1381612823666170707165319
- [6] Aguilar-Arnal L, Sassone-Corsi P. Chromatin landscape and circadian dynamics: Spatial and temporal organization of clock transcription. *PNAS*. 2015;112:6863-6870. DOI: 10.1073/pnas.1411264111
- [7] Peng H, Zhu Y, Goldberg J, Vaccarino V, Zhao J. DNA methylation of five core circadian genes jointly contributes to glucose metabolism: A gene-set analysis in monozygotic twins. *Frontiers in Genetics*. 2019;10:329. DOI: 10.3389/fgene.2019.00329
- [8] Duez H, Staels B, Ne Duez ÈÈHL, Staels B. Circadian control of epigenetic modifications modulates metabolism. *Circulation Research*. 2011;109:353-355. DOI: 10.1161/RES.0b013e31822be420
- [9] Grimaldi B, Nakahata Y, Kaluzova M, Masubuchi S, Sassone-Corsi P. Chromatin remodeling, metabolism and circadian clocks: The interplay of CLOCK and SIRT1. *The International Journal of Biochemistry & Cell Biology*. 2009;41:81-86. DOI: 10.1016/j.biocel.2008.08.035
- [10] Hermida RC, Ayala DE, Fernández JR, Calvo C. Chronotherapy improves blood pressure control and reverts the nondipper pattern in patients with resistant hypertension chronotherapy in resistant hypertension. *Hypertension*. 2007;51:69-76. DOI: 10.1161/HYPERTENSIONAHA.107.096933
- [11] Martino TA, Sole MJ. Molecular time: An often overlooked dimension to cardiovascular disease. *Circulation Research*. 2009;105:1047-1061. DOI: 10.1161/CIRCRESAHA.109.206201
- [12] Sun C, Burgner DP, Ponsonby AL, Saffery R, Huang RC, Vuillermin PJ, et al. Effects of early-life environment and epigenetics on cardiovascular disease risk in children: Highlighting the role of twin studies. *Pediatric Research*. 2013;73:523-530. DOI: 10.1038/pr.2013.6
- [13] Voelter-Mahlknecht S. Epigenetic associations in relation to cardiovascular prevention and therapeutics. *Clinical Epigenetics*. 2016;8:1-17. DOI: 10.1186/s13148-016-0170-0
- [14] Prasher D, Greenway SC, Singh RB. The impact of epigenetics on cardiovascular disease. *Biochemistry and Cell Biology*. 2019;98:12-22. DOI: 10.1139/bcb-2019-0045
- [15] Kearney PM, Whelton M, Reynolds K, Muntner P, Whelton PK, He J. Global burden of hypertension: Analysis of worldwide data. *Lancet*. 2005;365:217-223. DOI: 10.1016/s0140-6736(05)17741-1

- [16] Webster ALH, Yan MSC, Marsden PA. Epigenetics and cardiovascular disease. *The Canadian Journal of Cardiology*. 2013;**29**:46-57. DOI: 10.1016/j.cjca.2012.10.023
- [17] Arpón A, Milagro FI, Ramos-Lopez O, Mansego ML, Santos JL, Riezu-Boj JI, et al. Epigenome-wide association study in peripheral white blood cells involving insulin resistance. *Scientific Reports*. 2019;**9**:2445. DOI: 10.1038/s41598-019-38980-2
- [18] Campi3n J, Milagro FI, Mart3nez JA. Individuality and epigenetics in obesity. *Obesity Reviews*. 2009;**10**:383-392. DOI: 10.1111/j.1467-789X.2009.00595.x
- [19] Sahar S, Sassone-Corsi P. Circadian rhythms and memory formation: Regulation by chromatin remodeling. *Frontiers in Molecular Neuroscience*. 2012;**5**:1-4. DOI: 10.3389/fnmol.2012.00037
- [20] Kim JK, Samaranayake M, Pradhan S. Epigenetic mechanisms in mammals. *Cellular and Molecular Life Sciences*. 2009;**66**:596-612. DOI: 10.1007/s00018-008-8432-4
- [21] Borrelli E, Nestler EJ, Allis CD, Sassone-Corsi P. Decoding the epigenetic language of neuronal plasticity. *Neuron*. 2008;**60**:961-974. DOI: 10.1016/j.neuron.2008.10.012
- [22] Milagro FI, G3mez-Abell3n P, Campi3n J, Mart3nez JA, Ordov3s JM, Garaulet M. CLOCK, PER2 and BMAL1 DNA methylation: Association with obesity and metabolic syndrome characteristics and monounsaturated fat intake. *Chronobiology International*. 2012;**29**:1180-1194. DOI: 10.3109/07420528.2012.719967
- [23] Bannister AJ, Kouzarides T. Regulation of chromatin by histone modifications. *Cell Research*. 2011;**21**:381-395. DOI: 10.1038/cr.2011.22
- [24] Ripperger JA, Schibler U. Rhythmic CLOCK-BMAL1 binding to multiple E-box motifs drives circadian Dbp transcription and chromatin transitions. *Nature Genetics*. 2006;**38**:369-374. DOI: 10.1038/ng1738
- [25] Rossetto D, Avvakumov N, C3t3 J. Histone phosphorylation: A chromatin modification involved in diverse nuclear events. *Epigenetics*. 2012;**7**:1098-1108. DOI: 10.4161/epi.21975
- [26] Wilkinson KA, Henley JM. Mechanisms, regulation and consequences of protein SUMOylation. *The Biochemical Journal*. 2010;**428**:133-145. DOI: 10.1042/BJ20100158
- [27] Liu X, Liu S. Role of microRNAs in the pathogenesis of diabetic cardiomyopathy. *Biomedical Reports*. 2017;**6**:140-145. DOI: 10.3892/br.2017.841
- [28] Feng D, Lazar MA. Clocks, metabolism, and the epigenome. *Molecular Cell*. 2012;**47**:158-167. DOI: 10.1016/j.molcel.2012.06.026
- [29] Xie T, Gorenjak VG, Stathopoulou M, Dad3 S, Marouli E, Masson C, et al. Epigenome-wide association study (EWAS) of blood lipids in healthy population from STANISLAS family study (SFS). *International Journal of Molecular Sciences*. 2019;**20**:1014. DOI: 10.3390/ijms20051014
- [30] Kelleher FC, Rao A, Maguire A. Circadian molecular clocks and cancer. *Cancer Letters*. 2014;**342**:9-18. DOI: 10.1016/j.canlet.2013.09.040
- [31] Škrlec I, Marić S, Včev A. Myocardial infarction and circadian rhythm. In: Tsipis A, editor. *Visions Cardiomyocyte—Fundam. Concepts Hear. Life Dis*. 1st ed. London: IntechOpen; 2019. DOI: 10.5772/INTECHOPEN.83393
- [32] Škrlec I. Circadian rhythm and myocardial infarction. *Medicina*

Fluminensis. 2019;55:32-42. DOI: 10.21860/medflum2019\_216321

[33] Antypa N, Mandelli L, Nearchou FA, Vaiopoulos C, Stefanis CN, Serretti A, et al. The 311T/C polymorphism interacts with stressful life events to influence patterns of sleep in females. *Chronobiology International*. 2012;29:891-897. DOI: 10.3109/07420528.2012.699380

[34] Škrlec I, Milić J, Heffer M, Wagner J, Peterlin B. Circadian clock genes and circadian phenotypes in patients with myocardial infarction. *Advances in Medical Sciences*. 2019;64:224-229. DOI: 10.1016/j.advms.2018.12.003

[35] Hardeland R. Melatonin and chromatin. *Melatonin Research*. 2019;2:67-93. DOI: 10.32794/mr11250012

[36] Langmesser S, Tallone T, Bordon A, Rusconi S, Albrecht U. Interaction of circadian clock proteins PER2 and CRY with BMAL1 and CLOCK. *BMC Molecular Biology*. 2008;9:41. DOI: 10.1186/1471-2199-9-41

[37] Ohdo S, Koyanagi S, Matsunaga N. Chronopharmacological strategies: Intra- and inter-individual variability of molecular clock. *Advanced Drug Delivery Reviews*. 2010;62:885-897. DOI: 10.1016/j.addr.2010.04.005

[38] Partch CL, Green CB, Takahashi JS. Molecular architecture of the mammalian circadian clock. *Trends in Cell Biology*. 2014;24:90-99. DOI: 10.1016/j.tcb.2013.07.002

[39] Takeda N, Maemura K. Cardiovascular disease, chronopharmacotherapy, and the molecular clock. *Advanced Drug Delivery Reviews*. 2010;62:956-966. DOI: 10.1016/j.addr.2010.04.011

[40] Virag JAI, Lust RM. Circadian influences on myocardial infarction.

*Frontiers in Physiology*. 2014;5:422. DOI: 10.3389/fphys.2014.00422

[41] Kumar J, P Challet E, Kalsbeek A. Circadian rhythms in glucose and lipid metabolism in nocturnal and diurnal mammals. *Molecular and Cellular Endocrinology*. 2015;418:74-88. DOI: 10.1016/j.mce.2015.01.024

[42] Leu H-B, Chung C-M, Lin S-J, Chiang K-M, Yang H-C, Ho H-Y, et al. Association of circadian genes with diurnal blood pressure changes and non-dipper essential hypertension: A genetic association with young-onset hypertension. *Hypertension Research*. 2015;38:155-162. DOI: 10.1038/hr.2014.152

[43] Rudic RD, McNamara P, Curtis A-M, Boston RC, Panda S, Hogenesch JB, et al. BMAL1 and CLOCK, two essential components of the circadian clock, are involved in glucose homeostasis. *PLoS Biology*. 2004;2:e377. DOI: 10.1371/journal.pbio.0020377

[44] Curtis AM, Seo SB, Westgate EJ, Rudic RD, Smyth EM, Chakravarti D, et al. Histone acetyltransferase-dependent chromatin remodeling and the vascular clock. *The Journal of Biological Chemistry*. 2004;279:7091-7097. DOI: 10.1074/jbc.M311973200

[45] Turan N, Katari S, Coutifaris C, Sapienza C. Explaining inter-individual variability in phenotype: Is epigenetics up to the challenge? *Epigenetics*. 2010;5:16-19. DOI: 10.4161/epi.5.1.10557

[46] Aslibekyan S, Claas SA, Arnett DK. Epigenetics in cardiovascular disease. *Translating Epigenetics to the Clinic*. 2017;1:135-157. DOI: 10.1016/B978-0-12-800802-7.00006-X

[47] Turunen MP, Aavik E, Ylä-Herttuala S. Epigenetics and atherosclerosis. *Biochimica et Biophysica Acta*. 1790;2009:886-891. DOI: 10.1016/j.bbagen.2009.02.008

- [48] Gluckman PD, Hanson MA, Buklijas T, Low FM, Beedle AS. Epigenetic mechanisms that underpin metabolic and cardiovascular diseases. *Nature Reviews. Endocrinology*. 2009;**5**:401-408. DOI: 10.1038/nrendo.2009.102
- [49] Libby P, Okamoto Y, Rocha VZ, Folco E. Inflammation in atherosclerosis: Transition from theory to practice. *Circulation Journal*. 2010;**74**:213-220. DOI: 10.1253/circj.CJ-09-0706
- [50] Zhang L, Lin QL, Lu L, Yang CC, Li YL, Sun FL, et al. Tissue-specific modification of clock methylation in aging mice. *European Review for Medical and Pharmacological Sciences*. 2013;**17**:1874-1880
- [51] Bhatti P, Zhang Y, Song X, Makar KW, Sather CL, Kelsey KT, et al. Nightshift work and genome-wide DNA methylation. *Chronobiology International*. 2015;**32**:103-112. DOI: 10.3109/07420528.2014.956362
- [52] Nakahata Y, Grimaldi B, Sahar S, Hirayama J, Sassone-Corsi P. Signaling to the circadian clock: Plasticity by chromatin remodeling. *Current Opinion in Cell Biology*. 2007;**19**:230-237. DOI: 10.1016/j.ceb.2007.02.016
- [53] Westerman K, Sebastiani P, Jacques P, Liu S, DeMeo D, Ordovás JM. DNA methylation modules associate with incident cardiovascular disease and cumulative risk factor exposure. *Clinical Epigenetics*. 2019;**11**:142. DOI: 10.1186/s13148-019-0705-2
- [54] Azzi A, Dallmann R, Casserly A, Rehrauer H, Patrignani A, Maier B, et al. Circadian behavior is light-reprogrammed by plastic DNA methylation. *Nature Neuroscience*. 2014;**17**:377-382. DOI: 10.1038/nn.3651
- [55] Feinberg AP. Phenotypic plasticity and the epigenetics of human disease. *Nature*. 2007;**447**:433-440. DOI: 10.1038/nature05919
- [56] Leproult R, Holmbäck U, Van Cauter E. Circadian misalignment augments markers of insulin resistance and inflammation, independently of sleep loss. *Diabetes*. 2014;**63**:1860-1869. DOI: 10.2337/db13-1546
- [57] Gemma C, Sookoian S, Dieuzeide G, García SI, Gianotti TF, González CD, et al. Methylation of TFAM gene promoter in peripheral white blood cells is associated with insulin resistance in adolescents. *Molecular Genetics and Metabolism*. 2010;**100**:83-87. DOI: 10.1016/j.ymgme.2010.02.004
- [58] Zhu Y, Stevens RG, Hoffman AE, Tjonneland A, Vogel UB, Zheng T, et al. Epigenetic impact of long-term shiftwork: Pilot evidence from circadian genes and whole-genome methylation analysis. *Chronobiology International*. 2011;**28**:852-861. DOI: 10.3109/07420528.2011.618896
- [59] Ripperger JA, Meroz M. Perfect timing: Epigenetic regulation of the circadian clock. *FEBS Letters*. 2011;**585**:1406-1411. DOI: 10.1016/j.febslet.2011.04.047
- [60] Bozek K, Relógio A, Kielbasa SM, Heine M, Dame C, Kramer A, et al. Regulation of clock-controlled genes in mammals. *PLoS One*. 2009;**4**:e4882. DOI: 10.1371/journal.pone.0004882
- [61] Dong C, Yoon W, Goldschmidt-Clermont PJ. DNA methylation and atherosclerosis. *The Journal of Nutrition*. 2002;**132**:2406S-2409S. DOI: 10.1093/jn/132.8.2406s
- [62] Oka D, Yamashita S, Tomioka T, Nakanishi Y, Kato H, Kaminishi M, et al. The presence of aberrant DNA methylation in noncancerous esophageal mucosae in association with smoking history: A target for risk diagnosis and prevention of esophageal cancers.

Cancer. 2009;**115**:3412-3426. DOI: 10.1002/cncr.24394

[63] Samblas M, Milagro FI, Gómez-Abellán P, Martínez JA, Garaulet M. Methylation on the circadian gene BMAL1 is associated with the effects of a weight loss intervention on serum lipid levels. *Journal of Biological Rhythms*. 2016;**31**:308-317. DOI: 10.1177/0748730416629247

[64] Škrlec I, Milić J, Cilenšek I, Petrovič D, Wagner J, Peterlin B. Circadian clock genes and myocardial infarction in patients with type 2 diabetes mellitus. *Gene*. 2019;**701**:98-103. DOI: 10.1016/J.GENE.2019.03.038

[65] Škrlec I, Milić J, Heffer M, Peterlin B, Wagner J. Genetic variations in circadian rhythm genes and susceptibility for myocardial infarction. *Genetics and Molecular Biology*. 2018;**41**:403-409. DOI: 10.1590/1678-4685-gmb-2017-0147

[66] Sharma P, Kumar J, Garg G, Kumar A, Patowary A, Karthikeyan G, et al. Detection of altered global DNA methylation in coronary artery disease patients. *DNA and Cell Biology*. 2008;**27**:357-365. DOI: 10.1089/dna.2007.0694

[67] Jiang D, Sun M, You L, Lu K, Gao L, Hu C, et al. DNA methylation and hydroxymethylation are associated with the degree of coronary atherosclerosis in elderly patients with coronary heart disease. *Life Sciences*. 2019;**224**:241-248. DOI: 10.1016/j.lfs.2019.03.021

[68] Jiang YZ, Jiménez JM, Ou K, McCormick ME, Di Zhang L, Davies PF. Hemodynamic disturbed flow induces differential DNA methylation of endothelial Kruppel-like factor 4 promoter in vitro and in vivo. *Circulation Research*. 2014;**115**:32-43. DOI: 10.1161/CIRCRESAHA.115.303883

[69] Ward-Caviness CK, Agha G, Chen BH, Pfeiffer L, Wilson R, Wolf P, et al. Analysis of repeated leukocyte DNA methylation assessments reveals persistent epigenetic alterations after an incident myocardial infarction. *Clinical Epigenetics*. 2018;**10**:161. DOI: 10.1186/s13148-018-0588-7

[70] Baccarelli A, Wright R, Bollati V, Litonjua A, Zanobetti A, Tarantini L, et al. Ischemic heart disease and stroke in relation to blood DNA methylation. *Epidemiology*. 2010;**21**:819-828. DOI: 10.1097/EDE.0b013e3181f20457

[71] Nakatochi M, Ichihara S, Yamamoto K, Naruse K, Yokota S, Asano H, et al. Epigenome-wide association of myocardial infarction with DNA methylation sites at loci related to cardiovascular disease. *Clinical Epigenetics*. 2017;**9**:54. DOI: 10.1186/s13148-017-0353-3

[72] Castro R, Rivera I, Blom HJ, Jakobs C, Tavares de Almeida I. Homocysteine metabolism, hyperhomocysteinaemia and vascular disease: An overview. *Journal of Inherited Metabolic Disease*. 2006;**29**:3-20. DOI: 10.1007/s10545-006-0106-5

[73] Gómez-Uriz AM, Goyenechea E, Campión J, De Arce A, Martinez MT, Puchau B, et al. Epigenetic patterns of two gene promoters (TNF- $\alpha$  and PON) in stroke considering obesity condition and dietary intake. *Journal of Physiology and Biochemistry*. 2014;**70**:603-614. DOI: 10.1007/s13105-014-0316-5

[74] Jones MJ, Goodman SJ, Kobor MS. DNA methylation and healthy human aging. *Aging Cell*. 2015;**14**:924-932. DOI: 10.1111/accel.12349

[75] Sahar S, Sassone-Corsi P. The epigenetic language of circadian clocks. *Handbook of Experimental Pharmacology*. 2013:29-44. DOI: 10.1007/978-3-642-25950-0\_2

- [76] Cantó C, Auwerx J. Caloric restriction, SIRT1 and longevity. *Trends in Endocrinology and Metabolism*. 2009;**20**:325-331. DOI: 10.1016/j.tem.2009.03.008
- [77] Doi M, Hirayama J, Sassone-Corsi P. Circadian regulator CLOCK is a histone acetyltransferase. *Cell*. 2006;**125**:497-508. DOI: 10.1016/j.cell.2006.03.033
- [78] Etchegaray JP, Lee C, Wade PA, Reppert SM. Rhythmic histone acetylation underlies transcription in the mammalian circadian clock. *Nature*. 2003;**421**:177-182. DOI: 10.1038/nature01314
- [79] Masri S, Zocchi L, Katada S, Mora E, Sassone-Corsi P. The circadian clock transcriptional complex: Metabolic feedback intersects with epigenetic control. *Annals of the New York Academy of Sciences*. 2012;**1264**:103-109. DOI: 10.1111/j.1749-6632.2012.06649.x
- [80] Masri S, Sassone-Corsi P. The circadian clock: A framework linking metabolism, epigenetics and neuronal function. *Nature Reviews Neuroscience*. 2013;**14**:69-75. DOI: 10.1038/nrn3393
- [81] Naruse Y, Oh-hashii K, Iijima N, Naruse M, Yoshioka H, Tanaka M. Circadian and light-induced transcription of clock gene *Per1* depends on histone acetylation and deacetylation. *Molecular and Cellular Biology*. 2004;**24**:6278-6287. DOI: 10.1128/mcb.24.14.6278-6287.2004
- [82] Haigis MC, Guarente LP. Mammalian sirtuins—Emerging roles in physiology, aging, and calorie restriction. *Genes & Development*. 2006;**20**:2913-2921. DOI: 10.1101/gad.1467506
- [83] Nakahata Y, Kaluzova M, Grimaldi B, Sahar S, Hirayama J, Chen D, et al. The NAD<sup>+</sup>-dependent deacetylase SIRT1 modulates CLOCK-mediated chromatin remodeling and circadian control. *Cell*. 2008;**134**:329-340. DOI: 10.1016/j.cell.2008.07.002
- [84] Nakahata Y, Sahar S, Astarita G, Kaluzova M, Sassone-Corsi P. Circadian control of the NAD<sup>+</sup> salvage pathway by CLOCK-SIRT1. *Science*. 2009;**324**:654-657. DOI: 10.1126/science.1170803
- [85] Ramsey KM, Yoshino J, Brace CS, Abrassart D, Kobayashi Y, Marcheva B, et al. Circadian clock feedback cycle through NAMPT-mediated NAD<sup>+</sup> biosynthesis. *Science*. 2009;**324**:651-654. DOI: 10.1126/science.1171641
- [86] Asher G, Gatfield D, Stratmann M, Reinke H, Dibner C, Kreppel F, et al. SIRT1 regulates circadian clock gene expression through PER2 deacetylation. *Cell*. 2008;**134**:317-328. DOI: 10.1016/j.cell.2008.06.050
- [87] Longo VD, Kennedy BK. Sirtuins in aging and age-related disease. *Cell*. 2006;**126**:257-268. DOI: 10.1016/j.cell.2006.07.002
- [88] Zhang P, Yu Y, Qin Y, Zhou Y, Tang R, Wang Q, et al. Alterations to the microbiota-colon-brain axis in high-fat-diet-induced obese mice compared to diet-resistant mice. *The Journal of Nutritional Biochemistry*. 2019;**65**:54-65. DOI: 10.1016/j.jnutbio.2018.08.016
- [89] Feng D, Liu T, Sun Z, Bugge A, Mullican SE, Alenghat T, et al. A circadian rhythm orchestrated by histone deacetylase 3 controls hepatic lipid metabolism. *Science*. 2011;**331**:1315-1319. DOI: 10.1126/science.1198125
- [90] Fontaine C, Dubois G, Duguay Y, Helledie T, Vu-Dac N, Gervois P, et al. The orphan nuclear receptor rev-Erb $\alpha$  is a peroxisome proliferator-activated receptor (PPAR)  $\gamma$  target gene and promotes PPAR $\gamma$ -induced adipocyte

- differentiation. *The Journal of Biological Chemistry*. 2003;**278**:37672-37680. DOI: 10.1074/jbc.M304664200
- [91] Preitner N, Damiola F, Luis-Lopez-Molina L, Zakany J, Duboule D, Albrecht U, et al. The orphan nuclear receptor REV-ERB $\alpha$  controls circadian transcription within the positive limb of the mammalian circadian oscillator. *Cell*. 2002;**110**:251-260. DOI: 10.1016/S0092-8674(02)00825-5
- [92] Akashi M, Takumi T. The orphan nuclear receptor ROR $\alpha$  regulates circadian transcription of the mammalian core-clock Bmal1. *Nature Structural & Molecular Biology*. 2005;**12**:441-448. DOI: 10.1038/nsmb925
- [93] Matsushima S, Sadoshima J. The role of sirtuins in cardiac disease. *American Journal of Physiology-Heart and Circulatory Physiology*. 2015;**309**:H1375-H1389. DOI: 10.1152/ajpheart.00053.2015
- [94] Williams SM, Golden-Mason L, Ferguson BS, Schuetze KB, Cavasin MA, Demos-Davies K, et al. Class I HDACs regulate angiotensin II-dependent cardiac fibrosis via fibroblasts and circulating fibrocytes. *Journal of Molecular and Cellular Cardiology*. 2014;**67**:112-125. DOI: 10.1016/j.yjmcc.2013.12.013
- [95] Wada TT, Araki Y, Sato K, Aizaki Y, Yokota K, Kim YT, et al. Aberrant histone acetylation contributes to elevated interleukin-6 production in rheumatoid arthritis synovial fibroblasts. *Biochemical and Biophysical Research Communications*. 2014;**444**:682-686. DOI: 10.1016/j.bbrc.2014.01.195
- [96] Wang Y, Miao X, Liu Y, Li F, Liu Q, Sun J, et al. Dysregulation of histone acetyltransferases and deacetylases in cardiovascular diseases. *Oxidative Medicine and Cellular Longevity*. 2014;**2014**:1-11. DOI: 10.1155/2014/641979
- [97] Xie M, Kong Y, Tan W, May H, Battiprolu PK, Pedrozo Z, et al. Histone deacetylase inhibition blunts ischemia/reperfusion injury by inducing cardiomyocyte autophagy. *Circulation*. 2014;**129**:1139-1151. DOI: 10.1161/CIRCULATIONAHA.113.002416
- [98] Pitchiaya S, Heinicke LA, Park JI, Cameron EL, Walter NG. Resolving subcellular miRNA trafficking and turnover at single-molecule resolution. *Cell Reports*. 2017;**19**:630-642. DOI: 10.1016/j.celrep.2017.03.075
- [99] Cheng HYM, Papp JW, Varlamova O, Dziema H, Russell B, Curfman JP, et al. microRNA modulation of circadian-clock period and entrainment. *Neuron*. 2007;**54**:813-829. DOI: 10.1016/j.neuron.2007.05.017
- [100] Ahlin F, Arfvidsson J, Vargas KG, Stojkovic S, Huber K, Wojta J. MicroRNAs as circulating biomarkers in acute coronary syndromes: A review. *Vascular Pharmacology*. 2016;**81**:15-21. DOI: 10.1016/j.vph.2016.04.001
- [101] Wagschal A, Najafi-Shoushtari SH, Wang L, Goedeke L, Sinha S, Delemos AS, et al. Genome-wide identification of microRNAs regulating cholesterol and triglyceride homeostasis. *Nature Medicine*. 2015;**21**:1290-1297. DOI: 10.1038/nm.3980
- [102] Goedeke L, Rotllan N, Canfrán-Duque A, Aranda JF, Ramírez CM, Araldi E, et al. MicroRNA-148a regulates LDL receptor and ABCA1 expression to control circulating lipoprotein levels. *Nature Medicine*. 2015;**21**:1280-1288. DOI: 10.1038/nm.3949
- [103] Tan X, Zhang P, Zhou L, Yin B, Pan H, Peng X. Clock-controlled mir-142-3p can target its activator, Bmal1. *BMC Molecular Biology*. 2012;**13**:27. DOI: 10.1186/1471-2199-13-27



- [104] Shende VR, Goldrick MM, Ramani S, Earnest DJ. Expression and rhythmic modulation of circulating microRNAs targeting the clock gene *Bmal1* in mice. *PLoS One*. 2011;**6**:e22586. DOI: 10.1371/journal.pone.0022586
- [105] Bartman CM, Oyama Y, Brodsky K, Khailova L, Walker L, Koeppen M, et al. Intense light-elicited upregulation of miR-21 facilitates glycolysis and cardioprotection through Per2-dependent mechanisms. *PLoS One*. 2017;**12**:e0176243. DOI: 10.1371/journal.pone.0176243
- [106] Eckle T, Hartmann K, Bonney S, Reithel S, Mittelbronn M, Walker LA, et al. Adora2b-elicited Per2 stabilization promotes a HIF-dependent metabolic switch crucial for myocardial adaptation to ischemia. *Nature Medicine*. 2012;**18**:774-782. DOI: 10.1038/nm.2728
- [107] Ji X, Takahashi R, Hiura Y, Hirokawa G, Fukushima Y, Iwai N. Plasma miR-208 as a biomarker of myocardial injury. *Clinical Chemistry*. 2009;**55**:1944-1949. DOI: 10.1373/clinchem.2009.125310
- [108] Brainard J, Gobel M, Bartels K, Scott B, Koeppen M, Eckle T. Circadian rhythms in anesthesia and critical care medicine: Potential importance of circadian disruptions. *Seminars in Cardiothoracic and Vascular Anesthesia*. 2015;**19**:49-60. DOI: 10.1177/1089253214553066
- [109] Brainard J, Gobel M, Scott B, Koeppen M, Eckle T. Health implications of disrupted circadian rhythms and the potential for daylight as therapy. *Anesthesiology*. 2015;**122**:1170-1175. DOI: 10.1097/ALN.0000000000000596
- [110] Ritchie H, Stothard E, Wright K. Entrainment of the human circadian clock to the light-dark cycle and its impact on patients in the ICU and nursing home settings. *Current Pharmaceutical Design*. 2015;**21**:3438-3442. DOI: 10.2174/1381612821666150706111155
- [111] Foster RG, Peirson SN, Wulff K, Winnebeck E, Vetter C, Roenneberg T. Sleep and circadian rhythm disruption in social jetlag and mental illness. *Progress in Molecular Biology and Translational Science*. 2013;**119**:325-346. DOI: 10.1016/B978-0-12-396971-2.00011-7
- [112] Depner CM, Stothard ER, Wright KP. Metabolic consequences of sleep and circadian disorders. *Current Diabetes Reports*. 2014;**14**. DOI: 10.1007/s11892-014-0507-z